

Bridging model and experiment in systems neuroscience with Cleo: the Closed-Loop, Electrophysiology, and Optophysiology simulation testbed



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Simulate complex ephys, opto, and 2P imaging experiments with ease

INTRODUCTION

TWO-PHOTON IMAGING AND PHOTOSTIMULATION

A microscope selects neurons in focus for imaging while a Gaussian ellipsoid models off-target 2P laser stimulation.



Signal and noise amplitude depend on the

PROTOTYPING NOVEL EXPERIMENTS

We reject a traveling wave via closed-loop inhibition in a rodent S1 model from Moldakarimov et al. [9] and simulate the effect of increased control latency:



Cleo turns a spiking neural network model into a virtual experiment—useful for experiment design, methods engineering, and model evaluation:



indicator, but are scaled by per-ROI expression level and pixels in focus:

$$\mathrm{SNR}_{i} = \frac{\Delta F / F_{01\mathrm{AP}}}{\sigma_{\mathrm{noise}}} \frac{\rho_{\mathrm{rel}_{i}}}{1/\sqrt{N_{i}}}.$$

Detailed models of spike-driven calcium and GECI dynamics underlie fluorescence measurements [5]:

$$\Delta F/F_{0} = \Delta F/F_{0\max} \left(\frac{1}{1 + (K_{d} / [CaB_{active}])^{n_{H}}} - \frac{1}{1 + (K_{d} / [Ca^{2+}]_{rest})^{n_{H}}} \right)$$

$$\frac{d\left[Ca^{2+}\right]}{dt} = -\gamma \frac{\left[Ca^{2+}\right] - \left[Ca^{2+}\right]_{rest}}{1 + \kappa_S + \kappa_B}$$

 $\Delta \left[\mathrm{Ca}^{2+} \right] \left(t_{\mathrm{spike}} \right) = \frac{\Delta \left[\mathrm{Ca}^{2+} \right]_T}{1 + \kappa_{\mathrm{S}} + \kappa_{\mathrm{B}}}.$

ELECTROPHYSIOLOGY

Virtual electrode contacts record spiking and LFP proxy signals [1, 2] from the network:



END-TO-END VALIDATION EXPERIMENTS

We reproduce LFP recordings of epileptiform activity from HPC model and data from Aussel et al. [6]. Ablations fail to do likewise:

e LFP proxy ablation **f**

By clamping PV firing rate to a range of targets, we find a threshold for disrupting plasticity in a V1 model by Wilmes and Clopath [10]:



 \tilde{O} 0.0

600 650

Target multi-uni

firing rate [Hz]

-00 650

600

550

500 -

450



Feedback control can evoke a reference SWR-like oscillation in a HPC model [11] without manual stimulus design or calibration. Cleo makes it easy to try different control strategies:

600 650

Target multi-unit

firing rate [Hz]



OPTOGENETICS

Light propagation and opsin kinetics models enable simulation of one-photon optogenetics [3]:



Cleo is able to replicate the all-optical control experiment of Rickgauer et al. [7], using alternate opsin and calcium indicators:



Cleo also simulates multi-site/opsin stimulation, including crosstalk from overlapping action spectra [4]:



Using a simple E/I network model, we replicate Newman et al. [8]'s bidirectional "optoclamp" of *in vitro* firing rate. We use both the original and an alternate opsin pair:



RELATED WORK

Cleo is part of a larger effort developing and applying closed-loop optogenetic control (CLOC) methods:



See https://cloctools.github.io and Willats et al. [12, 13, 14], Bolus et al. [15, 16].

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References

- [1] Alberto Mazzoni, Henrik Lindén, Hermann Cuntz, Anders Lansner, Stefano Panzeri, and Gaute T. Einevoll. Computing the Local Field Potential (LFP) from Integrate-and-Fire Network Models. *PLOS Computational Biology*, 11(12):e1004584, 2015. ISSN 1553-7358. doi: 10.1371/JOURNAL.PCBI.1004584. URL https://journals.plos.org/ploscompbiol/article?id= 10.1371/journal.pcbi.1004584.
- [2] Bartosz Telenczuk, Maria Telenczuk, and Alain Destexhe. A kernel-based method to calculate local field potentials from networks of spiking neurons. *Journal of Neuroscience Methods*, 344:108871, October 2020. ISSN 1872678X. doi: 10.1016/j.jneumeth.2020.108871.
- [3] Thomas J Foutz, Richard L Arlow, and Cameron C Mcintyre. Theoretical principles underlying optical stimulation of a channelrhodopsin-2 positive pyramidal neuron. J Neurophysiol, 107: 3235–3245, 2012. doi: 10.1152/jn.00501.2011.-Optogenetics. [4] Himanshu Bansal, Neha Gupta, and Sukhdev Roy. Theoretical Analysis of Low-power Bidirectional Optogenetic Control of High-frequency Neural Codes with Single Spike Resolution. Neuro*science*, 449:165–188, November 2020. ISSN 18737544. doi: 10.1016/j.neuroscience.2020.09.022. [5] Alexander Song, Jeff L. Gauthier, Jonathan W. Pillow, David W. Tank, and Adam S. Charles. Neural anatomy and optical microscopy (NAOMi) simulation for evaluating calcium imaging methods. *Journal of Neuroscience Methods*, 358:109173, July 2021. ISSN 0165-0270. doi: 10.1016/j. jneumeth.2021.109173. [6] Amélie Aussel, Radu Ranta, Olivier Aron, Sophie Colnat-Coulbois, Louise Maillard, and Laure Buhry. Cell to network computational model of the epileptic human hippocampus suggests specific roles of network and channel dysfunctions in the ictal and interictal oscillations. *Journal of* Computational Neuroscience, August 2022. ISSN 1573-6873. doi: 10.1007/s10827-022-00829-5. URL https://doi.org/10.1007/s10827-022-00829-5. [7] John Peter Rickgauer, Karl Deisseroth, and David W. Tank. Simultaneous cellular-resolution optical perturbation and imaging of place cell firing fields. Nature Neuroscience, 17(12):1816–1824, December 2014. ISSN 1546-1726. doi: 10.1038/nn.3866. URL https://www.nature.com/articles/

nn.3866.

- [8] Jonathan P. Newman, Ming Fai Fong, Daniel C. Millard, Clarissa J. Whitmire, Garrett B. Stanley, and Steve M. Potter. Optogenetic feedback control of neural activity. *eLife*, 2015. ISSN 2050084X. doi: 10.7554/eLife.07192.
- [9] Samat Moldakarimov, Maxim Bazhenov, Daniel E. Feldman, and Terrence J. Sejnowski. Structured networks support sparse traveling waves in rodent somatosensory cortex. *Proceedings of the National Academy of Sciences of the United States of America*, 115(20):5277–5282, May 2018. ISSN 10916490. doi: 10.1073/pnas.1710202115. URL https://pubmed.ncbi.nlm.nih.gov/29712831/.
 [10] Katharina Anna Wilmes and Claudia Clopath. Inhibitory microcircuits for top-down plasticity of sensory representations. *Nature Communications*, 10(1):5055, November 2019. ISSN 2041-1723. doi: 10.1038/s41467-019-12972-2.
- [11] Amélie Aussel, Laure Buhry, Louise Tyvaert, and Radu Ranta. A detailed anatomical and mathematical model of the hippocampal formation for the generation of sharp-wave ripples and theta-nested gamma oscillations. *Journal of Computational Neuroscience*, 45(3):207–221, December 2018. ISSN 1573-6873. doi: 10.1007/s10827-018-0704-x.
- [12] A A Willats, M R O'Shaughnessy, and C J Rozell. Closed-loop identifiability in neural circuits. *In prep*, 2024.
- [13] A A Willats, M F Bolus, K A Johnsen, G B Stanley, and C J Rozell. Cloc tools: A library of tools for closed-loop neuroscience. *In prep*, 2024.
- [14] A A Willats, M F Bolus, C J Whitmire, G B Stanley, and C J Rozell. State-aware control of switching neural dynamics. *In prep*, 2024.
- [15] M F Bolus, A A Willats, C J Whitmire, C J Rozell, and G B Stanley. Design strategies for dynamic closed-loop optogenetic neurocontrol in vivo. *Journal of Neural Engineering*, 15(2):026011, April 2018. ISSN 1741-2560. doi: 10.1088/1741-2552/aaa506.
- [16] M F Bolus, A A Willats, C J Rozell, and G B Stanley. State-space optimal feedback control of optogenetically driven neural activity. *Journal of neural engineering*, 18(3):036006, June 2021. doi: 10.1101/2020.06.25.171785.